

Concepts of Fever

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If asked to define fever, most physicians would offer a thermal definition, such as “fever is a temperature greater than. . . .” In offering their definition, many would ignore the importance of the anatomic site at which temperature measurements are taken, as well as the diurnal oscillations that characterize body temperature.¹ If queried about the history of clinical thermometry, few physicians could identify the source or explain the pertinacity of the belief that 98.6°F (37.0°C) has special meaning vis-à-vis normal body temperature. Fewer still could cite the origin of the thermometer or trace the evolution of modern concepts of clinical thermometry. Although many would have some knowledge of the fundamentals of thermoregulation and the role played by exogenous and endogenous pyrogens in the induction of fever, few would have more than a superficial knowledge of the broad biological activities of pyrogenic cytokines or know of the existence of an equally complex and important system of endogenous cryogens. A distinct minority would appreciate the obvious paradoxes inherent in an enlarging body of data concerned with the question of fever’s adaptive value. The present review considers many of these issues in the light of current data.

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The oldest known written reference to fever exists in Akkadian cuneiform inscriptions from the sixth century BC, which seem to have been derived from an ancient Sumerian pictogram of a flaming brazier that symbolized fever and the local warmth of inflammation.² Theoretical constructs concerned with the pathogenesis of fever did not emerge until several centuries later, when Hippocratic physicians proposed that body temperature, and physiologic harmony in general, involved a delicate balance among 4 corporeal humors—blood, phlegm, black bile, and yellow bile.³ Fever, it was believed, resulted from an excess of yellow bile, a concept consistent with the fact that many infections of that era were associated with fever and jaundice. During the Middle Ages, demonic possession was added to the list of mechanisms believed responsible for fever. By the 18th century, Harvey’s discovery of the circulation

of blood and the birth of microbiology led iatrophysicists and iatrochemists to hypothesize, alternatively, that body heat and fever result from friction associated with the flow of blood through the vascular system and from fermentation and putrefaction occurring in the blood and intestines.⁴ Ultimately, thanks to the work of the great French physiologist, Claude Bernard, the metabolic processes occurring within the body finally came to be recognized as the source of body heat. Subsequent work established that body temperature is tightly controlled within a narrow range by mechanisms regulating the rate at which such heat is allowed to dissipate from the body.

The origin of the practice of monitoring body temperature as an aid to diagnosis is shrouded in uncertainty. The oldest known references to devices used

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to measure temperature date to the first or second century BC, when Philo of Byzantium and Hero of Alexandria are believed to have invented several such devices.⁵ It is reasonably certain that Galileo manufactured a primitive (air) thermometer at about the time he assumed the chair in mathematics at Padua in 1592.⁶ However, thermometry was not fully assimilated into medical practice until 1868, when Carl Reinhold August Wunderlich published a magnum opus entitled *Das Verhalten der Eigenwärme in Krankheiten* (*The Course of Temperature in Diseases*).⁷

Through *Das Verhalten der Eigenwärme in Krankheiten*, Wunderlich gave 98.6°F (37.0°C) its special meaning for normal body temperature.⁸ He described diurnal variation of body temperature and, in the process, alerted clinicians to the fact that “normal body temperature” is actually a temperature range rather than a specific temperature. In an analysis of a series of clinical thermometric measurements, the size of which has never been equaled (estimated to have included some 1 million observations in as many as 25 000 subjects), Wunderlich established 100.4°F (38.0°C) as the upper limit of the normal range and, in so doing, proffered one of the first quantitative definitions of fever.

Despite the fact that Wunderlich's work was published more than a century ago and was based primarily on axillary measurements generally taken no more often than twice daily, it has survived almost verbatim in modern day concepts of clinical thermometry. Interestingly, recent tests conducted with one of Wunderlich's thermometers suggest that his instruments may have been calibrated by as much as 1.4°C to 2.2°C (2.6°F-4.0°F) higher than today's instruments.⁸ As a result, at least some of Wunderlich's cherished dictums about body temperature (eg, the special significance of 98.6°F [37.0°C]) have required revision.⁹

DEFINITIONS

Fever has been defined as “a state of elevated core temperature, which is often, but not necessarily, part of the

defensive responses of multicellular organisms (host) to the invasion of live (microorganisms) or inanimate matter recognized as pathogenic or alien by the host.”¹⁰ The febrile response (of which fever is a component) is a complex physiologic reaction to disease, involving a cytokine-mediated rise in core temperature, generation of acute phase reactants, and activation of numerous physiologic, endocrinologic, and immunologic systems. The rise in temperature during fever is to be distinguished from that occurring during episodes of hyperthermia. Unlike fever, hyperthermia involves an unregulated rise in body temperature in which pyrogenic cytokines are not directly involved and against which standard antipyretics are ineffective. It represents a failure of thermoregulatory homeostasis, in which there is uncontrolled heat production, inadequate heat dissipation, or defective hypothalamic thermoregulation.

In the clinical setting, fever is typically defined as a pyrogen-mediated rise in body temperature above the normal range. Although useful as a descriptor for the febrile patient, the definition ignores the fact that a rise in body temperature is but one component of this multifaceted response. This standard clinical definition is further flawed, because it implies that “body temperature” is a single entity, when in fact, it is a pastiche of many different temperatures, each representative of a particular body part and each varying throughout the day in response to the activities of daily living and the influence of endogenous diurnal rhythms.

THERMOREGULATION

Heat is derived from biochemical reactions occurring in all living cells.¹¹ At the mitochondrial level, energy derived from the catabolism of metabolites, such as glucose, is used in oxidative phosphorylation to convert adenosine diphosphate to adenosine triphosphate. At rest, more than half of the body's heat is generated as a result of the inefficiency of the biochemical processes that convert food energy into the free en-

ergy pool (eg, adenosine triphosphate). When no external work is performed, the remainder of the body's heat (approximately 45%) is derived from the internal work involved in maintaining the structural and functional integrity of the body (ie, the use and resynthesis of adenosine triphosphate). When external work is performed, a portion of the latter heat (up to 25%) is generated by skeletal muscle contractions.

In adult humans and most other large mammals, shivering is the primary means by which heat production is enhanced. Nonshivering thermogenesis is more important in smaller mammals, newborns (including humans), and cold-acclimated mammals.^{11,12} Although several tissues (eg, the heart, respiratory muscles, and adipose tissue) may be involved, brown adipose tissue has been associated most closely with nonshivering thermogenesis. This highly specialized form of adipose tissue located near the shoulder blades, neck, adrenal glands, and deep blood vessels is characterized by its brownish color, a profuse vascular system, and an abundance of mitochondria.^{11,13}

Heat, generated primarily by vital organs lying deep within the body core, is distributed throughout the body by the circulatory system. In response to input from the nervous system, the circulatory system determines the temperature of the various body parts and the rate at which heat is lost from body surfaces to the environment (via conduction, convection, radiation, and evaporation).¹⁴ In a warm environment or in response to an elevation in core temperature due to exercise, cutaneous blood flow increases so that heat is transported from the core to be dissipated at the skin surface. In anesthetized animals, although discrete hypothalamic warming increases such cutaneous blood flow, blood pressure is maintained because of a concomitant reduction in gastrointestinal blood flow.¹⁵ In a cold environment or in response to a decrease in core temperature, cutaneous blood flow normally decreases as a means of retaining heat within the body core.¹⁴

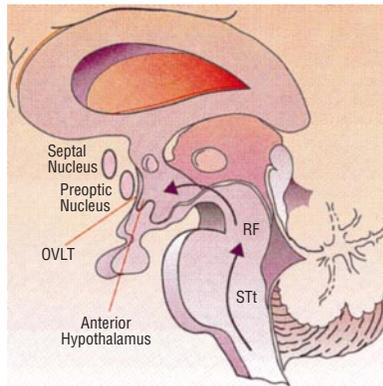


Figure 1. Sagittal view of the brain and upper spinal cord showing the multisynaptic pathway of skin and spinal thermoreceptors through the spinothalamic tract (STt) and reticular formation (RF) to the anterior hypothalamus, preoptic region, and the septum. OVLt indicates organum vasculosum laminae terminalis. Adapted from Mackowiak and Boulant.¹⁶

No single center within the nervous system controls body temperature. Rather, thermoregulation is a process that involves a continuum of neural structures and connections extending from the hypothalamus and limbic system through the lower brainstem and reticular formation to the spinal cord and sympathetic ganglia (**Figure 1**).¹⁶ Nevertheless, an area of the brain located in and near the rostral hypothalamus seems to have a pivotal role in the process of thermoregulation. Although generally referred to as the *preoptic region*, it actually includes the medial and lateral aspects of the preoptic area, anterior hypothalamus, and septum. Numerous studies extending more than 60 years have established that neurons located in this region are thermosensitive and exert at least partial control over physiologic and behavioral thermoregulatory responses.^{14,17}

Many, although not all, thermophysicologists believe that the temperature-sensitive preoptic region “regulates” body temperature by integrating thermal input signals from thermosensors in the skin and core areas (including the central nervous system).¹⁸ One of the more widely held theories is that such integration involves a designated thermal set point for the preoptic region that is maintained via a system of negative feedback. According to this theory, if the preoptic temperature rises above its set point for whatever reason (eg, dur-

ing exercise), heat loss responses are activated to lower body temperature and return the temperature of the preoptic region to the thermal set point (eg, 37.0°C).¹⁹ The thermal set point of a particular heat loss response is thus the maximum temperature tolerated by the preoptic region before the response is evoked. If, on the other hand, the preoptic temperature falls below its thermal set point (eg, as a result of cold exposure), various heat retention and heat production responses are activated to raise body temperature and with it the temperature of the preoptic region to its thermal set point. The thermal set point of a particular heat production response is thus the minimum temperature tolerated by the preoptic region before the response is evoked.

Although useful in explaining the elevation of the thermal set point that occurs during fever, the concept of a single central set point temperature is regarded by many thermal physiologists as oversimplified. At least some of these physiologists prefer to think of body temperature as regulated within a narrow range of temperatures by a composite set point of several thermosensitive areas and several different thermoregulatory responses.^{20,21}

A variety of endogenous substances and drugs seem to affect temperature regulation by altering the activity of hypothalamic neurons. Perhaps the best examples of such substances are the pyrogenic cytokines in the next section. These are released by phagocytic leukocytes in response to a wide array of stimuli and have the capacity to raise the thermoregulatory center’s thermal set point. Whether they cross the blood-brain barrier to do so^{22,23} or act by causing the release of other mediators (eg, prostaglandin E₂) in circumventricular organs, such as the organum vasculosum laminae terminalis²² is, as yet, uncertain. Whatever the precise endogenous mediators of fever, their primary effect seems to be to decrease the firing rate of preoptic warm-sensitive neurons, leading to activation of responses designed to decrease heat loss and increase heat production.

ENDOGENOUS PYROGENS

Pyrogens traditionally have been divided into 2 general categories: those that originate outside the body (exogenous pyrogens) and those derived from host cells (endogenous pyrogens). Exogenous pyrogens are, for the most part, microbes, toxins, or other products of microbial origin,²⁴ whereas endogenous pyrogens are host cell-derived (pyrogenic) cytokines that are the principal central mediators of the febrile response. According to current concepts, exogenous pyrogens, regardless of physicochemical structure, initiate fever by inducing host cells (primarily macrophages) to produce endogenous pyrogens. Such concepts notwithstanding, certain endogenous molecules also have the capacity to induce endogenous pyrogens. These include, among others, antigen-antibody complexes in the presence of complement,^{25,26} certain androgenic corticosteroid metabolites,²⁷⁻²⁹ inflammatory bile acids,³⁰ complement,³¹ and various lymphocyte-derived molecules.³²⁻³⁴

Complete understanding of the function of individual pyrogenic cytokines has been hampered by the fact that one cytokine often influences expression of other cytokines and/or their receptors and also may induce more distal comediators of cytokine-related bioactivities (eg, prostaglandins and platelet-aggregating factor).³⁵ In short, cytokines function within a complex regulatory network in which information is conveyed to cells by combinations and, perhaps, sequences of a host of cytokines and other hormones.³⁶ Like the words of human communication, individual cytokines are basic units of information. On occasion, a single cytokine, like a single word, may communicate a complete message. More often, however, complete messages received by cells probably resemble sentences, in which combinations and sequences of cytokines convey information. Because of such interactions, it has been difficult to ascertain the direct *in vivo* bioactivities of particular cytokines. Nevertheless, several cytokines have in common the capacity to induce fe-

Table 1. Biological Characteristics of the Principal Pyrogenic Cytokines*

Pyrogenic Cytokine	Aliases	Cell Source	Expression		Effect on Other Pyrogenic Cytokines	Biological Activities				
			Up-regulated by	Down-regulated by						
IL-1	Endogenous pyrogen Leukocyte endogenous pyrogen Lymphocyte-activating factor Mononuclear factor Catabolin Osteoclast-activating factor Hematopoietin-1 Melanoma growth inhibition factor Tumor inhibitory factor-2	Astrocytes	TNF	IL-4	↑ IL-6 ↑ TNF ↑ IL-1	IL-2 and IL-2R induction Thymocyte costimulation Fibroblast activation Induce acute phase response T-cell activation Costimulation of B-cell proliferation and differentiation Augment CTL, LAK induction Induce endothelial adhesion molecules Enhance phagocyte microbial killing Accelerate wound healing				
		Endothelial cells	IFN- γ	IL-6						
		Keratinocytes	GM-CSF	IL-10						
		Monocytes	Zymosan	TGF- β						
		Macrophages	LPS	Corticosteroids						
		Dendrites	IL-1	PGE ₂						
		Fibroblasts	C5a	Retinoic acid						
			Leukotrienes PMA							
TNF- α	Cachectin	Monocytes	Bacteria	Corticosteroids	↑ TNF ↑ IL-1 ↑ IL-6	Septic shock Enhance phagocyte microbial killing Tumor necrosis Cachexia Anorexia Endothelial and epithelial MHC, adhesion molecule induction Osteoclast activation B-cell differentiation CTL induction				
		Macrophages	Viruses	Cyclosporine						
		Eosinophils	Fungi	PGE ₂						
		Neutrophils	Protozoa	IL-4						
		Lymphocytes	LPS	IL-6						
		Astrocytes	Staph TSST1	IL-10						
		Endothelial cells	IL-1	TGF- β						
		Mast cells	IL-2	Vitamin D ₃						
		Kupffer cells	TNF							
		NK cells	IFNs							
		Certain tumors	GM-CSF PAF Substance P Anti-TCR Tumor cells PMA							
		IL-6	Interferon beta-2 B-cell stimulatory factor-2 Hybridoma or plasmacytoma growth factor Hepatocyte-stimulating factor Cytotoxic T-cell differentiation factor Macrophage granulocyte-inducing factor 2A	Monocytes			LPS	Corticosteroids	↓ TNF ↓ IL-1	B-cell growth, differentiation, and IgG synthesis Myeloma proliferation CTL induction Acute phase response Thymocyte costimulation Weak antiviral activity Megakaryocyte maturation Neuronal differentiation Enhance IL-3-dependent stem cell proliferation
				Macrophages			IL-1	Estrogens		
				Lymphocytes			TNF			
Fibroblasts	IFN- β Calcium ionophore									
Endothelial cells	Mitogenic lectin and									
Epithelial cells										
Keratinocytes	PMA									
Bone marrow stroma	Viruses									
Certain tumors										
IFN- γ	Type II interferon Immune interferon	T cells	Mitogenic lectins	Corticosteroids	↑ TNF ↑ IL-1	Macrophage priming Antiviral activity Enhance TNF activity MHC induction Enhance NK activity Enhance endothelial ICAM-1 expression Inhibit IL-4-induced B-cell responses B-cell differentiation and IgG2a secretion				
		NK cells	IL-1 IL-2	Cyclosporine Vitamin D ₃						

*IL indicates interleukin; TNF, tumor necrosis factor; IFN, interferon; GM-CSF, granulocyte-macrophage colony-stimulating factor; LPS, lipopolysaccharide; C5a, complement component C5a; PMA, phorbol myristate acetate; TGF- β , transforming growth factor β ; PGE₂, prostaglandin E₂; CTL, cytotoxic T lymphocytes; LAK, lymphocyte-activated killer; NK, natural killer; Staph TSST1, staphylococcal toxic shock syndrome toxin-1; PAF, platelet-activating factor; TCR, T-cell receptor; MHC, major histocompatibility complex; and ICAM, intercellular adhesion molecule. An upward arrow indicates enhanced expression; a downward arrow, reduced expression. Adapted from Hasday and Goldblum cited in Mackowiak et al.³⁵

ver. Based on this characteristic, they have been codified as so-called pyrogenic cytokines.

The list of currently recognized pyrogenic cytokines includes, among others, interleukin (IL)-1 (IL-1 α and IL-1 β), tumor necrosis factor α (TNF- α), IL-6, and interferon gamma (IFN- γ) (Table 1).³⁷⁻⁴⁵ Even among these

few cytokines, complex relationships exist, with certain members up-regulating expression of other members or their receptors under certain conditions and down-regulating them under other conditions.³⁵ The 4 pyrogenic cytokines have monomeric molecular weights that range from 17 to 30 kd. Undetectable under basal conditions in

healthy subjects, they are produced by many different tissues in response to appropriate stimuli. Once released, pyrogenic cytokines have short intravascular half-lives. They are pleiotropic: they interact with receptors present on many different host cells. They are active in picomolar quantities, induce maximal cellular responses

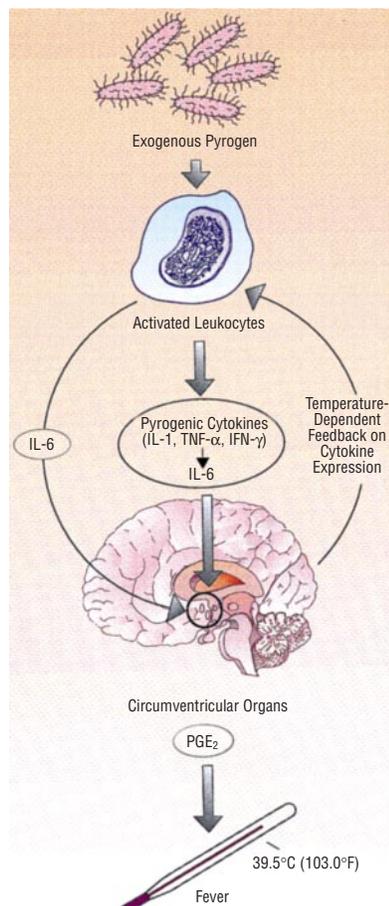


Figure 2. Hypothetical model for the febrile response. IL indicates interleukin; TNF, tumor necrosis factor; IFN, interferon; and PGE₂, prostaglandin E₂.

even at low receptor occupancy, and exert local (autocrine or paracrine) and systemic (endocrine) effects.³⁵

It has been theorized, although not proved, that interaction between pyrogenic cytokines and their receptors in the preoptic region of the anterior hypothalamus activates phospholipase A₂, liberating plasma membrane arachidonic acid as substrate for the cyclooxygenase pathway. Some cytokines might do so by increasing cyclooxygenase expression directly, causing liberation of the arachidonate metabolite, prostaglandin E₂. Because this small lipid molecule easily diffuses across the blood-brain barrier, it might be the local mediator that actually activates thermosensitive neurons. Although not discussed in this article, recent studies indicate that thermal information involved in the febrile response also might be transmitted from the periphery to the thermo-

Table 2. The Acute Phase Proteins (ACPs)*

Positive ACPs†
C-reactive protein
Serum amyloid A
Haptoglobin
α ₁ -Acid glycoprotein
α ₁ -Protease inhibitor
Fibrinogen
Ceruloplasmin
Complement (C3 and C4)
C1 esterase inhibitor
C4b binding protein
α ₂ -Macroglobulin
Ferritin
Phospholipase A ₂
Plasminogen activator inhibitor-1
Fibronectin
Hemopexin
Pancreatic secretory trypsin inhibitor
Inter-α protease inhibitor
Manose binding protein
Negative ACPs‡
Albumin
Transthyretin
Transferrin
α ₂ -HS glycoprotein

*Adapted from Kushner and Rzewnicki.³⁶

†Proteins exhibiting increased plasma concentrations during the acute phase response.

‡Proteins exhibiting decreased plasma concentrations during the acute phase response.

regulatory center via peripheral nerves.⁴⁶

Extensive work with pyrogenic cytokines during the last 2 decades has provided a hypothetical model for the febrile response (**Figure 2**). Nevertheless, our understanding of this process remains incomplete and largely speculative. As indicated, several issues remain unresolved: (1) whether circulating cytokines cross the blood-brain barrier or have to be produced within the central nervous system to activate thermosensitive neurons; (2) whether each of the pyrogenic cytokines is capable of raising the thermoregulatory set point independently or must exert this effect through some common pathway (eg, IL-6, as suggested by Dinarello²⁴; **Figure 2**); (3) whether prostaglandin E₂ or other local mediators are a sine qua non of the febrile response; (4) what determines the magnitude of expression of individual cytokines in response to various stimuli; and (5) how the upper limit of the febrile range is set.³⁵

Tumor necrosis factor α and IL-1 have pivotal roles during the induction phase of the febrile response⁴⁷ but also are expressed throughout the response.⁴⁸ A small but growing body of data suggests that temperatures near the upper end of the febrile range influence production of such cytokines.⁴⁹⁻⁶⁰ However, such effects are highly dependent on experimental conditions.

THE ACUTE PHASE RESPONSE

As noted, a cytokine-mediated rise in core temperature is but one of many features of the febrile response. Numerous other physiologic reactions, collectively referred to as the *acute phase response*, are mediated by members of the same group of pyrogenic cytokines that activate the thermal response of fever.³⁶ Such reactions include somnolence, anorexia, changes in plasma protein synthesis, and altered synthesis of hormones such as corticotropin-releasing hormone, glucagon, insulin, corticotropin, hydrocortisone, adrenal catecholamines, growth hormone, thyrotropin, thyroxine, aldosterone, and arginine vasopressin. Inhibition of bone formation, negative nitrogen balance, gluconeogenesis, and altered lipid metabolism also are seen during the acute phase response, as are decreased serum concentrations of zinc and iron and increased serum concentrations of copper. Hematologic alterations⁶¹ include leukocytosis, thrombocytosis, and decreased erythropoiesis (resulting in an "anemia of chronic inflammation"⁶²). Stimuli capable of inducing an acute phase response include bacterial and, to a lesser extent, viral infection, trauma, malignant neoplasms, burns, tissue infarction, immunologically mediated and crystal-induced inflammatory states, strenuous exercise,⁶³ and childbirth. Recent data also suggest that major depression,⁶⁴ schizophrenia,⁶⁵ and psychological stress⁶⁶ are capable of inducing an acute phase response.

Traditionally, the phrase *acute phase response* has been used to denote changes in plasma concentrations of a number of secretory proteins derived from hepatocytes. *Acute*

phase proteins, of which there are many (**Table 2**),³⁶ exhibit increased synthesis (positive acute phase proteins) or decreased synthesis (negative acute phase proteins) during the acute phase response.

Many of the acute phase proteins are believed to modulate inflammation and tissue repair.⁶⁷ A major function of C-reactive protein (CRP), for example, is presumed to involve binding of phosphocholine on pathogenic microorganisms, as well as phospholipid constituents on damaged or necrotic host cells. Through such binding, CRP might activate the complement system and promote phagocyte adherence, thereby initiating the process by which pathogenic microbes or necrotic cells are eliminated from the host. Such activities are most likely potentiated by CRP-induced production of inflammatory cytokines⁶⁸ and tissue factor⁶⁹ by monocytes. Nevertheless, the ultimate function of CRP is uncertain; several *in vivo* studies have shown it to have anti-inflammatory properties.⁷⁰⁻⁷²

The other major human acute phase protein, serum amyloid A, recently has been reported to potentiate adhesiveness and chemotaxis of phagocytic cells and lymphocytes.⁷³ There also is evidence that macrophages bear specific binding sites for serum amyloid A, that serum amyloid A-rich, high-density lipoproteins mediate transfer of cholesterol to macrophages at sites of inflammation,⁷⁴ and that serum amyloid A enhances low-density lipoprotein oxidation in arterial walls.⁷⁵

Complement components, many of which are acute phase reactants, modulate chemotaxis, opsonization, vascular permeability, and vascular dilatation and have cytotoxic effects as well.³⁶ Haptoglobin, hemopexin, and ceruloplasmin all are antioxidants. It is, therefore, reasonable to assume that, like the antiproteases, α_1 -antichymotrypsin and C1 esterase inhibitor, they have important roles in modulating inflammation. However, the functional capacity of such proteins is broad.

Although closely associated with fever, the acute phase response is not an invariable compo-

nent of the febrile response.³⁶ Some febrile patients (eg, those with certain viral infections) have normal blood levels of CRP. Moreover, patients with elevated blood levels of CRP are not always febrile. The acute phase response, like the febrile response, is a complex response consisting of numerous integrated but separately regulated components. The particular components expressed in response to a given disease process more than likely reflects the specific cytokines induced by the disease.

ENDOGENOUS CRYOGENS

Hippocrates maintained that "Heat is the immortal substance of life endowed with intelligence. . . . However, heat must also be refrigerated by respiration and kept within bounds if the source or principle of life is to persist; for if refrigeration is not provided, the heat will consume itself."⁷⁶ Modern day clinicians also generally subscribe to the notion that the febrile range has an upper limit,¹ but do not agree on a precise temperature defining this limit. The lack of a consensus in this regard is understandable, owing to the fact that "body" temperature profiles exhibit considerable individual, anatomic, and diurnal variability. For this reason, the upper limit of the febrile range cannot be defined as a single temperature applicable to all body sites of all people at all times during the day. Nevertheless, the febrile response is a regulated physiologic response, in which temperature is maintained within certain carefully controlled limits, the upper limit of which almost never exceeds 41.0°C, regardless of the cause of the fever or site at which temperature measurements are taken.⁷⁷ The physiologic necessity of this upper limit is supported by considerable experimental data demonstrating adverse physiologic effects of core temperatures greater than 41.0°C or 42.0°C.¹⁶

The mechanisms regulating fever's upper limit have yet to be fully defined. They could lie with the intrinsic properties of the neurons themselves or involve the release of endogenous antipyretic substances that antagonize the effects of pyro-

gens on thermosensitive neurons. For the former possibility, plots of the firing rates of neurons coordinating thermoregulatory responses and heat production tend to converge at 42.0°C (**Figure 3**).¹⁶ At this temperature, the long-term or extended firing rates of warm-sensitive neurons reach their zenith and cannot be increased further in response to higher temperatures. Similarly, the firing rates of cold-sensitive neurons reach their nadir at 42.0°C and cannot decrease further even if temperature increases further. Thus, regardless of pyrogen concentration, thermosensitive neurons seem to be incapable of providing additional thermoregulatory signals once the temperature reaches 42.0°C.

These same thermosensitive neurons are influenced by a variety of endogenous substances, at least some of which seem to function as endogenous cryogens.¹⁶ Studies by numerous investigators using a variety of animal models have established that arginine vasopressin is present in the fibers and terminals of the ventral septal area of the hypothalamus, is released into the ventral septal area during fever, and reduces fever via its action at type 1 vasopressin receptors when introduced into the ventral septal area and, when inhibited, prolongs fever.⁷⁸⁻⁸⁰

α -Melanocyte-stimulating hormone (α -MSH) is another neuropeptide exhibiting endogenous antipyretic activity.⁸¹ Unlike some other antipyretic peptides, α -MSH has not been identified in fibers projecting into the ventral septal area.⁸² It does, nevertheless, reduce pyrogen-induced fever when administered to experimental animals in doses below those having an effect on afebrile body temperature.⁸³⁻⁸⁷ When given centrally, α -MSH is more than 25 000 times more potent as an antipyretic than acetaminophen.⁸¹ Repeated central administration of α -MSH does not induce tolerance to its antipyretic effect.⁸⁸ In addition, injection of anti- α -MSH antiserum into the cerebral ventricles augments the febrile response of experimental animals to IL-1.⁸⁹

Numerous neurochemicals seem to have the capacity to influ-

ence hypothalamic control of body temperature. Because some lower body temperature even in the absence of fever, they are more appropriately termed *hypothermic* agents than *antipyretic* agents. In some of the earliest work in this area, Feldberg and Meyers⁹⁰ observed that intracerebroventricular injections of epinephrine and norepinephrine in cats cause a fall in body temperature, whereas injections of serotonin cause temperature to rise. Based on these observations, they proposed that regulation of body temperature involves a balance between the release of catecholamines (inducing heat loss) and serotonin (activating heat production) in the anterior hypothalamus. More recent data, including those considered in the present article, suggest that the basis of set-point determination by the thermoregulatory network is considerably more complex.⁹¹

Glucocorticoids and their inducers (corticotropin-releasing hormone and corticotropin) inhibit synthesis of pyrogenic cytokines such as IL-6 and TNF- α .⁹²⁻⁹⁴ Through such effects, they are believed to exert inhibitory feedback on lipopolysaccharide (LPS)-induced fever.⁹⁵ Lipocortin 1, a putative mediator of glucocorticoid function, also has been shown to inhibit the pyrogenic actions of IL-1 and IFN.⁹⁶ Corticotropin-releasing hormone injected into the third ventricle of experimental animals produces similar antipyretic effects.⁹⁷

Thyroliberin,⁹⁸ gastric inhibitory polypeptide,⁹⁹ neuropeptide Y,¹⁰⁰ and bombesin,¹⁰¹ likewise, exhibit cryogenic properties under appropriate conditions. Of these, bombesin is probably the most potent, because it consistently produces hypothermia associated with changes in heat dissipation and heat production when injected into the preoptic area/anterior hypothalamus of conscious goats and rabbits.¹⁰¹⁻¹⁰³ Bombesin is believed to exert its hypothermic effect by increasing the temperature sensitivity of warm-sensitive neurons.¹⁰²

Pyrogenic cytokines, the mediators of the febrile response, might themselves have a direct role in determining fever's upper limit. There

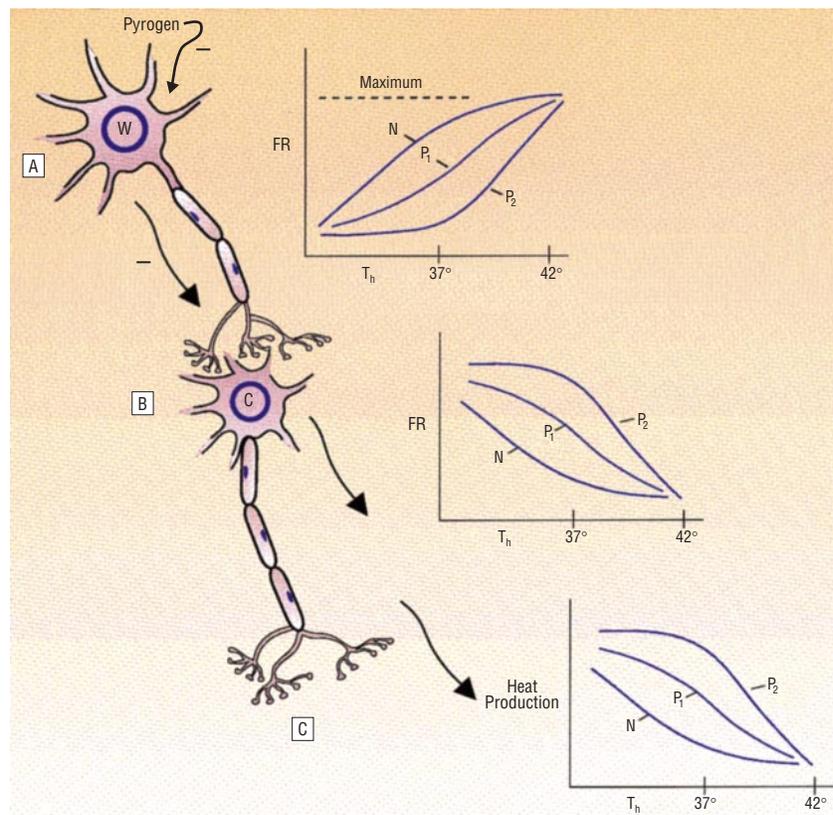


Figure 3. Model showing responses (A and B) of neuronal firing rates (FR) in the preoptic region and anterior hypothalamus and whole-body metabolic heat production (C) during changes in hypothalamic temperature (T_h). Thermosensitivity is reflected by the slope of each plot. The letters inside the cells indicate a warm-sensitive (w) neuron and a cold-sensitive (c) neuron. With increases in T_h , warm-sensitive neurons raise their FRs, and heat production decreases. Pyrogens inhibit (-) the FRs of warm-sensitive neurons, thereby resulting in accelerated FRs of cold-sensitive neurons and increased heat production. The plots show FR and heat production responses during normal conditions in the absence of pyrogens (N) and in the presence of low concentrations (P_1) and high concentrations (P_2) of pyrogens. The temperatures are given in degrees Celsius. Adapted from Mackowiak and Boulant.¹⁶

is, for instance, experimental evidence indicating that under certain conditions TNF- α lowers, rather than raises, body temperature.^{104,105} Thus, it is possible that, at certain concentrations or in the appropriate physiologic milieu (eg, at 41.0°-42.0°C), pyrogenic cytokines function paradoxically as endogenous cryogens.

A growing body of literature indicates that release of pyrogenic cytokines, such as IL-1, is followed by increased shedding of soluble receptors for such cytokines, which function as endogenous inhibitors of these pyrogens.¹⁰⁶ In the case of IL-1, a 22- to 25-kd molecule identified in supernatants of human monocytes blocks binding of IL-1 to its receptors.¹⁰⁷ The IL-1 receptor antagonist is structurally related to IL-1 α and IL-1 β ¹⁰⁸ and binds to type I and type II receptors on various target cells without inducing a specific bio-

logical response.^{109,110} Shedding of soluble circulating receptors of TNF- α that bind to circulating TNF- α and thereby inhibit binding to cell-associated receptors also has been described.¹¹¹⁻¹¹⁵ The precise biological function of such circulating receptor antagonists and soluble receptors is unknown. However, it is possible, that 1 function is to serve as a natural braking system for the febrile response.

RISK-BENEFIT CONSIDERATIONS

Questions about fever's risk-benefit quotient have generated considerable controversy in recent years.¹¹⁶ The controversy arises because of substantial data indicating potentiating and inhibitory effects of the response on resistance to infection. As a result, there is no consensus about the appropriate clinical

situations (if any) in which fever or its mediators should be suppressed.

Data illustrating fever's beneficial effects originate from several sources. Studies of the phylogeny of fever have shown the response to be widespread within the animal kingdom.¹¹⁷ With few exceptions, mammals, reptiles, amphibians, and fish, as well as several invertebrate species, have been shown to manifest fever in response to challenge with microorganisms or other known pyrogens. This fact has been viewed as some of the strongest evidence that fever is an adaptive response, based on the argument that the metabolically expensive increase in body temperature that accompanies the febrile response would not have evolved and been so faithfully preserved within the animal kingdom unless fever had some net benefit to the host.

Further evidence of fever's beneficial effects can be found in numerous studies demonstrating enhanced resistance of animals to infection with increases in body temperature within the physiologic range.¹¹⁷ In classic studies involving experimental infection of the reptile *Dipsosaurus dorsalis* with *Aeromonas hydrophila*, Kluger et al¹¹⁸ and Bernheim and Kluger¹¹⁹ demonstrated a direct correlation between body temperature and survival. Bernheim and Kluger¹¹⁹ also showed that suppression of the febrile response with sodium salicylate results in a substantial increase in mortality. Covert and Reynolds¹²⁰ corroborated these findings in an experimental model involving goldfish.

In mammalian experimental models, increasing body temperature by artificial means has been reported to enhance resistance of mice to herpes simplex virus,¹²¹ poliovirus,¹²² coxsackie B virus,¹²³ rabies virus,¹²⁴ and *Cryptococcus neoformans*,¹²⁵ but to decrease resistance to *Streptococcus pneumoniae*.¹²⁶ Increased resistance of rabbits to *S pneumoniae*¹²⁷ and *C neoformans*,¹²⁸ dogs to herpesvirus,¹²⁹ piglets to gastroenteritis virus,¹³⁰ and ferrets to influenza virus¹³¹ also has been observed after induction of artificial fever. Unfortunately, because rais-

ing body temperature by artificial means does not duplicate the physiologic alterations that occur during fever in homeotherms (and, indeed, entails a number of opposite physiologic responses¹³²), data obtained using mammalian experimental models have been less convincing than those obtained using reptiles or fish.

Clinical data supporting an adaptive role for fever have accumulated slowly. Like animal data, clinical data include evidence of beneficial effects of fever and adverse effects of antipyretics on the outcome of infections. In a retrospective analysis of 218 patients with gram-negative bacteremia, Bryant et al¹³³ reported a positive correlation between maximum temperature on the day of bacteremia and survival. A similar relationship has been observed in patients with polymicrobial sepsis and mild (but not severe) underlying diseases.¹³⁴ In an examination of factors influencing the prognosis of spontaneous bacterial peritonitis, Weinstein et al¹³⁵ identified a positive correlation between a temperature reading of more than 38°C and survival.

It has been reported that children with chicken pox who are treated with acetaminophen have a longer time to total crusting of lesions than placebo-treated control subjects.¹³⁶ Stanley et al¹³⁷ reported that adults infected with rhinovirus exhibit more nasal viral shedding when they receive aspirin than when given placebo. Furthermore, Graham and colleagues¹³⁸ reported a trend toward longer duration of rhinovirus shedding in association with antipyretic therapy and showed that the use of aspirin or acetaminophen is associated with suppression of the serum neutralizing antibody response and with increased nasal symptoms and signs. These data, like those reviewed in the preceding paragraph, are subject to several interpretations and do not prove a causal relationship between fever and improved prognosis during infection. Nevertheless, they are consistent with such a relationship, and, when considered in concert with the phylogeny of the febrile response and the animal data summarized herein, they constitute strong cir-

cumstantial evidence that fever is an adaptive response in most situations.

Whereas the foregoing studies focused on the relationship between elevation of core temperature and the outcome of infection, others have considered the endogenous mediators of the febrile response. In such studies, all 4 of the major pyrogenic cytokines have been shown to have immune-potentiating capabilities that might theoretically enhance resistance to infection (Table 1).¹³⁹ In vitro and in vivo studies of these cytokines have provided evidence of a protective effect of IFN, TNF- α , and/or IL-1 against *Plasmodium* organisms,¹⁴⁰⁻¹⁴² *Toxoplasma gondii*,¹⁴³ *Leishmania major*,¹⁴⁴ *Trypanosoma cruzi*,¹⁴⁵ and *Cryptosporidium* organisms.¹⁴⁶

Several recent reports also have shown enhancement of resistance to viral¹⁴⁷⁻¹⁴⁹ and bacterial^{150,151} infections by pyrogenic cytokines. Treatment of healthy and granulocytopenic animals with IL-1 has been shown to prevent death in some gram-positive and gram-negative bacterial infections.¹⁵¹ However, IL-1 is effective only if administered an appreciable time (eg, 24 hours) before initiation of infections having rapidly fatal courses. In less acute infections, IL-1 administration can be delayed until shortly after the infectious challenge. Such observations suggest that the physiologic effects of fever that enhance resistance to infection might be limited to localized infections or systemic infections of only mild to moderate severity.

The potential of the febrile response for harm is reflected in a recent flurry of reports suggesting that IL-1, TNF- α , IL-6, and IFN mediate the physiological abnormalities of certain infections. Although proof of an adverse effect of fever on the clinical outcome of these infections has yet to be established, the implication is that if pyrogenic cytokines contribute to the pathophysiologic burden of infections, the mediators themselves and the febrile response are potentially deleterious.

The most persuasive evidence derives from studies of gram-negative bacterial sepsis.¹⁵² It has long been suspected that bacterial LPS has a pivotal role in the syndrome. Puri-

fied LPS induces a spectrum of physiological abnormalities that are similar to those occurring in patients with gram-negative bacterial sepsis. In experimental animals, challenge with LPS causes TNF- α and IL-1 to be released into the bloodstream coincident with the appearance of signs of sepsis.¹⁵³ Furthermore, patients with the septic syndrome have detectable levels of circulatory TNF- α , IL-1, and IL-6 independent of culture-documented infection, and such levels correlate inversely with survival.¹⁵⁴ Interleukin 1, alone or in combination with other cytokines, induces many of the same physiological abnormalities (eg, fever, hypoglycemia, shock, and death) seen after administration of purified LPS.¹⁵⁵ In a murine experimental model for septic shock, IFN administered before or as long as 4 hours after LPS challenge increases mortality, whereas pretreatment with anti-IFN antibody substantially reduces mortality.¹⁵⁶ In several recent studies, the adverse effects of gram-negative bacterial sepsis, LPS injections, or both have been attenuated by pretreating experimental animals with IL-1 antagonists^{157,158} and monoclonal antibodies directed against TNF- α .^{159,160} Furthermore, animals rendered tolerant to TNF- α by repeated injections of the recombinant cytokine are protected against the hypotension, hypothermia, and lethality of gram-negative bacterial sepsis.¹⁶¹

Together, these observations have led to a growing conviction that pyrogenic cytokines are central mediators of the clinical and humoral manifestations of gram-negative bacterial sepsis and have generated intense interest, although little progress,¹⁶² in the clinical application of antagonists of such cytokines. Similar data suggest that pyrogenic cytokines might mediate at least some of the systemic and local manifestations of sepsis due to gram-positive bacteria,^{153,163,164} AIDS,¹⁶⁵ spirochetal infections,^{166,167} meningitis,¹⁶⁸ the adult respiratory distress syndrome,^{165,169} suppurative arthritis,¹⁷⁰ and mycobacteriosis.¹⁷¹

CONCLUSIONS

To fully appreciate the clinical implications of fever, one must take a

broad view that encompasses the febrile response in its entirety. Fever is mediated by a host of cytokines that not only cause the body's thermoregulatory set point to rise, but also simultaneously stimulate production of a panoply of acute phase reactants (although, apparently not invariably) and activate numerous metabolic, endocrinologic, and immunologic systems. For these reasons, fever cannot be equated with hyperthermia. More important, experimental models of "fever" in which body temperature is elevated by external means or by agents that markedly increase heat production by uncoupling oxidative phosphorylation must be recognized as having limited value in the study of this physiologic response. Only if one views fever from the perspective of its relationship with the febrile response, can one begin to explain the apparent paradox inherent in reports demonstrating beneficial effects of therapy with pyrogenic cytokines and their antagonists and, through such understanding, take maximum advantage of the response to alleviate the burden of human disease.

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REFERENCES

- Mackowiak PA, Wasserman SS. Physicians' perceptions regarding body temperature in health and disease. *South Med J*. 1995;88:934-938.
- Majno G. *The Healing Hand: Man and Wound in the Ancient World*. Cambridge, Mass: Harvard University Press; 1975:57.
- Galen. Opera omnia. In: Siegel RE, ed. *Galen's System of Physiology and Medicine*. Vol 11. New York, NY: Karger; 1968.
- Atkins E. Fever: its history, cause and function. *Yale J Biol Med*. 1982;55:283-287.
- Berger RL, Clem TR, Harden VA, Mangum BW. Historical development and newer means of temperature measurements in biochemistry. *Methods Biochem Anal*. 1984;269-331.
- Bolton HC. *Evolution of the Thermometer 1592-*

1743. Easton, Pa: Chemical Publishing Co; 1900: 18, 98.
- Wunderlich CRA. *Das Verhalten der Eigenwärme in Krankheiten*. Leipzig, Germany: Otto Wigard; 1868.
- Mackowiak PA, Worden G. Carl Reinhold August Wunderlich and the evolution of clinical thermometry. *Clin Infect Dis*. 1994;18:458-467.
- Mackowiak PA, Wasserman SS, Levine MM. A critical appraisal of 98.6°F: the upper limit of the normal body temperature, and other legacies of Carl Reinhold August Wunderlich. *JAMA*. 1992; 268:1578-1580.
- IUPS Thermal Commission. Glossary of terms for thermal physiology: second edition. *Pflugers Arch*. 1987;410:567-587.
- Boulant JA. Thermoregulation. In Mackowiak PA, ed. *Fever: Basic Mechanisms and Management*. 2nd ed. Philadelphia, PA: Lippincott-Raven Publishers; 1997:35-58.
- Bruck K. Heat balance and the regulation of body temperature. In: Schmidt RF, Thews G, eds. *Human Physiology*. Berlin, Germany: Springer-Verlag; 1983:531-547.
- Stanier MW, Mount LE, Bligh J. *Energy Balance and Temperature Regulation*. Cambridge, England: Cambridge University Press; 1984.
- Boulant JA. Hypothalamic control of thermoregulation: neurophysiological basis. In: Morgane PJ, Pankepp J, eds. *Handbook of the Hypothalamus*. Vol 3, part A. New York, NY: Marcel Dekker Inc; 1980:1-82.
- Schonung W, Wagner H, Jessen C, Simon E. Differentiation of cutaneous and intestinal blood flow during hypothalamic heating and cooling in anesthetized dogs. *Pflugers Arch*. 1971;328:145-154.
- Mackowiak PA, Boulant JA. Fever's glass ceiling. *Clin Infect Dis*. 1996;22:525-536.
- Boulant JA. Hypothalamic neurons regulating body temperature. In: Fregly MJ, Blatteis CM, eds. *APS Handbook of Physiology*. New York, NY: Oxford University Press; 1996:105-126.
- Hammel HT, Jackson DC, Stolwijk JAJ, Hardy JD, Stromme SB. Temperature regulation by hypothalamic proportional control with an adjustable set point. *J Appl Physiol*. 1963;18:1146-1154.
- Hammel HT. Neurons and temperature regulation. In: Yamamoto WS, Brobeck JR, eds. *Physiological Controls and Regulations*. Philadelphia, Pa: WB Saunders Co; 1965:71-97.
- Sawka MN, Wenger CB. Physiological responses to acute exercise-heat stress. In: Pandolf KB, Sawka MN, Gonzalez RR, eds. *Human Performance Physiology and Environmental Medicine at Terrestrial Extremes*. Indianapolis, Ind: Benchmark Press; 1988:97-151.
- Bligh J. *Temperature Regulation in Mammals and Other Vertebrates*. Amsterdam, the Netherlands: North Holland; 1973.
- Stitt JT. Prostaglandin E as the mediator of the febrile response. *Yale J Biol Med*. 1986;59:137-149.
- Mitchell D, Laburn HP, Cooper KE, Hellon RF, Cranston WI, Townsend Y. Is prostaglandin E the neural mediator of the febrile response? the case against a proven obligatory role. *Yale J Biol Med*. 1986;59:159-168.
- Dinarello CA. Cytokines as endogenous pyrogens. In: Mackowiak PA, ed. *Fever: Basic Mechanisms and Management*. 2nd ed. Philadelphia, Pa: Lippincott-Raven Publishers; 1997:87-116.
- Mickenberg ID, Snyderman R, Root RK, Mergenhagen SE, Wolff SM. The relationship of

- complement consumption to immune fever. *J Immunol.* 1971;107:1466-1476.
26. Arend WP, Joslin FG, Massoni RJ. Effects of immune complexes on production by human monocytes of interleukin 1 or an interleukin 1 inhibitor. *J Immunol.* 1985;134:3868-3875.
 27. Dillard GM, Bodel P. Studies on steroid fever, II: pyrogenic and anti-pyrogenic activity in vitro of some endogenous steroids of man. *J Clin Invest.* 1970;49:2418-2426.
 28. Kappas A, Hellman L, Fukushima DK, Gallagher TF. The pyrogenic effect of etiocholanolone [letter]. *J Clin Endocrinol Metab.* 1957;17:451-453.
 29. Wolff SM, Kimball HR, Perry S, Root RK, Kappas A. The biological properties of etiocholanolone. *Ann Intern Med.* 1967;67:1268-1295.
 30. Bondy PK, Bodel P. Mechanism of action of pyrogenic and antipyretic steroids in vitro. In: Wolstenholme GEW, Birch J, eds. *Pyrogens and Fever.* Edinburgh, Scotland: Churchill Livingstone Inc; 1971;101-113.
 31. Goodman MG, Chenoweth DE, Weigle WO. Induction of interleukin 1 secretion and enhancement of humoral immunity by binding of human C5a to macrophage surface C5a receptors. *J Exp Med.* 1982;156:912-917.
 32. Atkins E, Feldman JD, Francis L, Hursh E. Studies on the mechanism of fever accompanying delayed hypersensitivity: the role of the sensitized lymphocyte. *J Exp Med.* 1972;135:1113-1132.
 33. Bernheim HA, Block LH, Francis L, Atkins E. Release of endogenous pyrogen-activating factor from concanavalin A-stimulated human lymphocytes. *J Exp Med.* 1980;152:1811-1816.
 34. Dinarello CA. Demonstration of a human pyrogen-inducing factor during mixed leukocyte reactions. *J Exp Med.* 1981;153:1215-1224.
 35. Mackowiak PA, Barlett JG, Borden EC, et al. Fever: recent advances and lingering dogma. *Clin Infect Dis.* 1997;25:119-138.
 36. Kushner I, Rzewnicki DL. The acute phase response. In: Mackowiak PA, ed. *Fever: Basic Mechanisms and Management.* 2nd ed. Philadelphia, Pa: Lippincott-Raven Publishers; 1997:165-176.
 37. Dinarello CA, Wolff SM. The role of interleukin-1 in disease. *N Engl J Med.* 1993;328:106-113.
 38. Dinarello C. The interleukin-1 family: 10 years of discovery. *FASEB J.* 1994;8:1314-1325.
 39. Dinarello C. Interleukin-1. *Adv Pharmacol.* 1994;25:21-51.
 40. Fiers W. Tumor necrosis factor: characterization at the molecular, cellular and in vivo level. *FEBS Lett.* 1991;285:199-212.
 41. Vassalli P. The pathophysiology of tumor necrosis factors. *Annu Rev Immunol.* 1992;10:411-452.
 42. Tracey K, Cerami A. Tumor necrosis factor: a pleiotropic cytokine and therapeutic target. *Annu Rev Med.* 1994;45:491-503.
 43. Brach M, Herrman F. Interleukin 6: presence and future. *Int J Clin Lab Res.* 1992;22:143-151.
 44. Lotz M. Interleukin-6. *Cancer Invest.* 1993;11:731-742.
 45. Jones T. Interleukin-6 an endocrine cytokine. *Clin Endocrinol.* 1994;40:703-713.
 46. Blatteis CM, Sehic E. Prostaglandin E₂: a putative fever mediator. In: Mackowiak PA, ed. *Fever: Basic Mechanisms and Management.* 2nd ed. Philadelphia, Pa: Lippincott-Raven Publishers; 1997:117-145.
 47. Kluger M. Fever: role of pyrogens and cryogens. *Physiol Rev.* 1991;71:93-127.
 48. Michie HR, Manague KR, Spriggs DR, et al. Detection of circulating tumor necrosis factor after endotoxin administration. *N Engl J Med.* 1988;318:1481-1486.
 49. Klostergaard J, Barta M, Tomasovic SP. Hyperthermic modulation of tumor necrosis factor-dependent monocyte/macrophage tumor cytotoxicity in vitro. *J Biol Res Mod.* 1989;8:262-277.
 50. Tomasovic SP, Klostergaard J. Hyperthermic modulation of macrophage-tumor cell interactions. *Cancer Metastasis Rev.* 1989;8:215-229.
 51. Fouqueray B, Philippe C, Amrani A, Perez J, Baud L. Heat shock prevents lipopolysaccharide-induced tumor necrosis factor- α synthesis by rat mononuclear phagocytes. *Eur J Immunol.* 1992;22:2983-2987.
 52. Snyder Y, Guthrie ML, Evans GF, Zuckerman SH. Transcriptional inhibition of endotoxin-induced monokine synthesis following heat shock in murine peritoneal macrophages. *J Leukoc Biol.* 1992;51:181-187.
 53. Valasco S, Tarlow M, Olsen K, et al. Temperature-dependent modulation of lipopolysaccharide-induced interleukin-1 β and tumor necrosis factor- α expression in cultured human astroglia by dexamethasone and indomethacin. *J Clin Invest.* 1991;87:1674-1680.
 54. Kappel M, Diamant M, Hansen MB, Klokner M, Pedersen PK. Effects of in vitro hyperthermia on the proliferative response of blood mononuclear cell subsets, and detection of interleukins 1 and 6, tumor necrosis factor-alpha and interferon-gamma. *Immunology.* 1991;73:304-308.
 55. Ensor JE, Wiener SM, McCrea KA, Viscardi RM, Crawford EK, Hasday JD. Differential effects of hyperthermia on macrophage interleukin-6 and tumor necrosis factor- α expression. *Am J Physiol Cell Physiol.* 1994;266:C967-C974.
 56. Ensor JE, Crawford EK, Hasday JD. Warming macrophages to febrile range destabilizes tumor necrosis factor- α mRNA without inducing heat shock. *Am J Physiol Cell Physiol.* 1995;269:C1140-C1146.
 57. Weiner S, Hasday JD. Temperature dependence of monocyte cytokine release. *FASEB J.* 1991;5:A626.
 58. Costa J, DeTolla L, Piper J, Ensor J, Hasday JD. Effects of febrile range temperature on cytokine expression in lipopolysaccharide-challenged mice. *FASEB J.* 1995;9:A960.
 59. Aderka D, Le J, Vilcek J. IL-6 inhibits lipopolysaccharide-induced tumor necrosis factor production in cultured human monocytes, U937 cells, and in mice. *J Immunol.* 1989;143:3517-3523.
 60. Cross AS, Sadoff JC, Kelly N, Bernton E, Gemski P. Pretreatment with recombinant murine tumor necrosis factor α /cachectin and murine interleukin 1 α protects mice from lethal bacterial infection. *J Exp Med.* 1989;169:2021-2027.
 61. Trey J, Kushner I. The acute phase response and the hematopoietic system: the role of cytokines. *Crit Rev Oncol Hematol.* 1995;21:1-18.
 62. Schilling RF. Anemia of chronic disease: a misnomer. *Ann Intern Med.* 1991;115:572-573.
 63. Ernst E, Saradeth T, Achhammer G. ω 3 fatty acids and acute-phase proteins. *Eur J Clin Invest.* 1991;21:77-82.
 64. Joyce PR, Hawes CR, Mulder RT, Sellman JD, Wilson DA, Boswell DR. Elevated levels of acute phase plasma proteins in major depression. *Biol Psychiatry.* 1992;32:1035-1041.
 65. Ganguli R, Yang Z, Shurin G, et al. Serum interleukin-6 concentration in schizophrenia: elevation associated with duration of illness. *Psychiatry Res.* 1994;51:1-10.
 66. LeMay LG, Vander AJ, Kluger MJ. The effects of psychological stress on plasma interleukin-6 activity in rats. *Physiol Behav.* 1990;47:957-961.
 67. Volanakis JE. Acute phase proteins. McCarty DJ, Koopman WJ, eds. *Arthritis and Allied Conditions: A Textbook of Rheumatology.* Malvern, Pa: Lea & Febiger; 1993:469-477.
 68. Ballou SP, Lozanski G. Induction of inflammatory cytokines release from cultured human monocytes by C-reactive protein. *Cytokine.* 1992;4:361-368.
 69. Cermak J, Key NS, Bach RR, Jacob HS, Vercellotti GM. C-reactive protein induces human peripheral blood monocytes to synthesize tissue factor. *Blood.* 1993;82:513-520.
 70. Tilg H, Vannier E, Vachine G, Dinarello CA, Mier JW. Anti-inflammatory properties of hepatic acute phase proteins: preferential induction of interleukin 1 (IL-1) receptor antagonist over IL-1 β synthesis by human peripheral blood mononuclear cells. *J Exp Med.* 1993;178:1629-1636.
 71. Dobrinich R, Spagnuolo PJ. Binding of C-reactive protein to human neutrophils: inhibition of respiratory burst activity. *Arthritis Rheum.* 1991;34:1031-1038.
 72. Ahmed N, Thorley R, Xia D, Samols D, Webster RO. Transgenic mice expressing rabbit C-reactive protein exhibit diminished chemotactic factor-induced alveolitis. *Am J Respir Crit Care Med.* In press.
 73. Xu L, Badolato R, Murphy WJ, et al. A novel biologic function of serum amyloid A-induction of T lymphocyte migration and adhesion. *J Immunol.* 1995;155:1184-1190.
 74. Kisilevsky R, Subrahmanyam L. Serum amyloid A changes high-density lipoprotein's cellular affinity. *Lab Invest.* 1992;66:778.
 75. Berliner JA, Navab M, Fogelman AM, et al. Atherosclerosis: basic mechanisms: oxidation, inflammation, and genetics. *Circulation.* 1995;91:2488-2496.
 76. May MT. *Galen on the Usefulness of the Parts of the Body.* Ithaca, NY: Cornell University Press; 1968:50-53.
 77. DuBois EF. Why are fever temperatures over 106°F rare? *Am J Med Sci.* 1949;217:361-368.
 78. Pittman QJ, Wilkinson MF. Central arginine vasopressin and endogenous antipyresis. *Can J Physiol Pharmacol.* 1992;70:786-790.
 79. Pittman QJ, Poulin P, Wilkinson MF. Role of neurohypophysial hormones in temperature regulation. *Ann N Y Acad Sci.* 1993;689:375-381.
 80. Kastning N. Criteria for establishing a physiological role for brain peptides: a case in point: the role of vasopressin in thermoregulation during fever and antipyresis. *Brain Res Rev.* 1989;14:143-153.
 81. Lipton JM. Disorders of temperature control. In: Rieder P, Kopp N, Pearson J, eds. *An Introduction to Neurotransmission in Health and Disease.* Oxford, England: Oxford University Press; 1990:119-123.
 82. Zeisberger E. The role of septal peptides in thermoregulation and fever. In: Blish J, Voigt K, eds. *Thermoreception and Temperature Regulation.* Berlin, Germany: Springer-Verlag; 1990:273-283.
 83. Glyn JR, Lipton JM. Hypothermic and antipyretic effects of centrally administered ACTH (-24) and α -melanotropin. *Peptides.* 1981;2:177-187.
 84. Glyn-Ballinger JR, Bernardini GL, Lipton JM. α -

- MSH injected into the septal region reduces fever in rabbits. *Peptides*. 1983;4:199-203.
85. Lipton JM, Whisenant JD, Gean JT. Hypothermia produced by peripheral and central injections of chlorpromazine in aged rabbits. *Brain Res Bull*. 1979;4:297-300.
 86. Murphy MT, Lipton JM. Peripheral administration of α -MSH reduces fever in older and younger rabbits. *Peptides*. 1982;13:775-779.
 87. Murphy MT, Richard DB, Lipton JM. Antipyretic potency of centrally administered α -melanocyte-stimulating hormone. *Science*. 1983; 221:192-193.
 88. Deeter LB, Martin LW, Lipton JM. Antipyretic effect of central alpha-MSH summates with that of acetaminophen or ibuprofen. *Brain Res Bull*. 1989;23:573-575.
 89. Shih ST, Lipton JM, McCann SM. Central administration of α -MSH antiserum augments fever in the rabbit. *Am J Physiol*. 1986;250:R803-R806.
 90. Feldberg W, Meyers RD. A new concept of temperature regulation by amines in the hypothalamus. *Nature*. 1963;200:1325.
 91. Blich J. Cells, cell-talk and mammalian homeothermy. In: Blich J, Voigt K, eds. *Thermoreception and Temperature Regulation*. Berlin, Germany: Springer-Verlag; 1990:163-173.
 92. Morrow LE, McClellan JL, Conn CA, Kluger MJ. Glucocorticoids alter fever and IL-6 responses to psychological stress and to lipopolysaccharide. *Am J Physiol*. 1993;225:R151-R156.
 93. Luedke CE, Cerami A. Interferon-gamma overcomes glucocorticoid suppression of cachectin/tumor necrosis factor biosynthesis by murine macrophages. *J Clin Invest*. 1990;86:1234-1240.
 94. Nakano T, Ohara O, Teraoka H, Arita H. Glucocorticoids suppress group II phospholipase A_2 production by blocking mRNA synthesis and post-transcriptional expression. *J Biol Chem*. 1990;265:12745-12748.
 95. Alexander DP, Bashore RA, Britton HG, Forsling MA. Maternal and fetal arginine vasopressin in the chronically catheterized sheep. *Biol Neonate*. 1974;25:242-248.
 96. Carey F, Forder M, Edge D, et al. Lipocortin 1 fragment modifies pyrogenic actions of cytokines in rats. *Am J Physiol*. 1990;259:R266-R269.
 97. Bernadini GL, Lipton JM, Clark WG. Intracerebroventricular and septal injections of arginine vasopressin are not antipyretic in the rabbit. *Peptides*. 1983;4:195-198.
 98. Riedel W. Role of thyroid-stimulating hormone (TSH) in endogenous antipyraxis and evidence of extrahypothalamic thyroid-stimulating neurons (TSN) in rabbits. *Pflügers Arch*. 1987;408 (suppl):R49.
 99. Bahendeka SK, Moor RE, Tomkin GH, Buchanan KD. Gastric inhibitory polypeptide, dietary-induced thermogenesis and obesity. *Can J Physiol Pharmacol*. 1987;65:1242-1247.
 100. Stanley BG, Leibowitz SF, Neuropeptide Y. Stimulation of feeding and drinking by injection into the paraventricular nucleus. *Life Sci*. 1984;35: 2635-2642.
 101. Jansky L, Vybiral S, Moravec J, et al. Neuropeptides and temperature regulation. *J Therm Biol*. 1986;11:79-83.
 102. Schmid H, Pierau Fr-K. Long-term modulation of hypothalamic neurons by neuropeptides. In: Blich J, Voigt K, eds. *Thermoregulation and Temperature Regulation*. Berlin, Germany: Springer-Verlag; 1990:53-63.
 103. Gale CC, McCreery BR. Mechanism of bombesin hypothermia. *Fed Proc*. 1979;38:997.
 104. Holt SJ, Grimble RF, York DA. Tumor necrosis factor- α and lymphotoxin have opposite effects on sympathetic efferent nerves to brown adipose tissue by direct action in the central nervous system. *Brain Res*. 1989;497:183-186.
 105. Shih ST, Khorram O, Lipton JM, McCann SM. Central administration of α -MSH antiserum augments fever in the rabbit. *Am J Physiol*. 1986; 250:R803-R806.
 106. Sivo J, Salkowski CA, Politis AD, Vogel SN. Differential regulation of LPS-induced IL-1 β and IL-1 receptor antagonist mRNA by IFN α and IFN γ in murine peritoneal macrophages. *J Endotoxin Res*. 1994;1:30-36.
 107. Seckinger P, Lowenthal JW, Williamson K, Dayer JM, MacDonald HR. A urine inhibitor of interleukin-1 activity that blocks ligand binding. *J Immunol*. 1987;139:1546-1549.
 108. Eisenberg SP, Brewer MT, Verderber E, Heimdal P, Thompson RC. Interleukin-1 receptor antagonist is a member of the interleukin-1 gene family: evolution of a cytokine control mechanism. *Proc Natl Acad Sci U S A*. 1991;88:5232-5236.
 109. Dripps DJ, Brandhuber BJ, Thompson RC, Eisenberg SP. Interleukin-1 (IL-1) receptor antagonist binds to the 80-kDa IL-receptor but does not initiate IL-1 signal transduction. *J Biol Chem*. 1991;266:10331-10336.
 110. Dripps DJ, Verderber E, Ng RK, Thompson RC, Eisenberg SP. Interleukin-1 receptor antagonist binds to the type II interleukin-1 receptor on B cells and neutrophils. *J Biol Chem*. 1991;266:20311-20315.
 111. Engelmann H, Aderka D, Rubinstein M, Rotman D, Wallach D. A tumor necrosis factor binding protein purified to homogeneity from human urine protects cells from tumor necrosis factor toxicity. *J Biol Chem*. 1989;264:11974-11980.
 112. Olsson I, Lantx M, Nilsson E, et al. Isolation and characterization of a tumor necrosis factor binding protein from urine. *Eur J Haematol*. 1989; 42:270-275.
 113. Engleman H, Novick D, Wallach D. Two tumor necrosis factor-binding proteins from human urine. *J Biol Chem*. 1990;265:1531-1536.
 114. Brockhuus M, Schoenfeld HJ, Schlaeger EJ, Hunziker W, Lesslauer W, Loetscher M. Identification of two types of tumor necrosis factor receptors on human cell lines by monoclonal antibodies. *Proc Natl Acad Sci U S A*. 1990;87: 3127-3131.
 115. Porteu F, Nathan C. Shedding of tumor necrosis factor receptors by activated human neutrophils. *J Exp Med*. 1990;172:599-607.
 116. Mackowiak PA. Fever: blessing or curse? a unifying hypothesis. *Ann Intern Med*. 1994;120: 1037-1040.
 117. Kluger MJ, Kozat W, Conn CA, Leon LR, Soszynski D. The adaptive value of fever. In: Mackowiak PA, ed. *Fever: Basic Mechanisms and Management*. Philadelphia, Pa: Lippincott-Raven Publishers; 1997:255-266.
 118. Kluger MJ, Ringler DH, Anver MR. Fever and survival. *Science*. 1975;188:166-168.
 119. Bernheim HA, Kluger MJ. Fever: Effect of drug-induced antipyraxis on survival. *Science*. 1976; 193:237-239.
 120. Covert JR, Reynolds WW. Survival value of fever in fish. *Nature*. 1977;267:43-45.
 121. Schmidt JR, Rasmussen AF Jr. The influence of environmental temperature on the course of experimental herpes simplex infection. *J Infect Dis*. 1960;107:356-360.
 122. Lwoff A. Factors influencing the evolution of viral diseases at the cellular level and in the organism. *Bacteriol Rev*. 1959;23:109-124.
 123. Walker DL, Boring WD. Factors influencing host-virus interactions, III: further studies on the alteration of coxsackie virus infection in adult mice by environmental temperature. *J Immunol*. 1958; 80:39-44.
 124. Bell JF, Moore GJ. Effects of high ambient temperature on various stages of rabies virus infection in mice. *Infect Immun*. 1974;10:510-515.
 125. Kuhn LR. Effect of elevated body temperature on cryptococcosis in mice. *Proc Soc Exp Biol Med*. 1949;71:341-343.
 126. Eiseman B, Mallette WG, Wotkins RS, Summers WB, Tong JL. Prolonged hypothermia in experimental pneumococcal peritonitis. *J Clin Invest*. 1956;35:940-946.
 127. Rich AR, McKee CM. The mechanism of a hitherto unexplained form of native immunity to the type III pneumococcus. *Bull Johns Hopkins Hosp*. 1936;59:171-207.
 128. Kuhn LR. Growth and viability of *Cryptococcus hominis* at mouse and rabbit body temperatures. *Proc Soc Exp Biol Med*. 1939;41:573-574.
 129. Carmichael LE, Barnes FD. Effect of temperature on growth of canine herpes virus in canine kidney cell and macrophage cultures. *J Infect Dis*. 1969;120:664-668.
 130. Furuchi S, Shimizu Y. Effect of ambient temperatures on multiplication of attenuated transmissible gastroenteritis virus in the bodies of newborn piglets. *Infect Immun*. 1976;13:990-992.
 131. Toms GL, Davies JA, Woodward CG, Sweet C, Smith H. The relation of pyrexia and nasal inflammatory response to virus levels in nasal washings of ferrets infected with influenza viruses of differing virulence. *Br J Exp Pathol*. 1977; 588:444-458.
 132. Greisman SE. Cardiovascular alterations during fever. In: Mackowiak PA, ed. *Fever: Basic Mechanisms and Management*. New York, NY: Raven Press; 1991:143-165.
 133. Bryant RE, Hood AF, Hood CE, Koenig MG. Factors affecting mortality of gram-negative rod bacteremia. *Arch Intern Med*. 1971;127:120-128.
 134. Mackowiak PA, Browne RH, Southern PM Jr, Smith JW. Polymicrobial sepsis: analysis of 184 cases using log linear models. *Am J Med Sci*. 1980;280:73-80.
 135. Weinstein MR, Iannini PB, Staton CW, Eichoff TC. Spontaneous bacterial peritonitis: a review of 28 cases with emphasis on improved survival and factors influencing prognosis. *Am J Med*. 1978;64:592-598.
 136. Dorn TF, DeAngelis C, Baumgardner RA, et al. Acetaminophen: more harm than good for chicken pox? *J Pediatr*. 1989;114:1045-1048.
 137. Stanley ED, Jackson GG, Panusarn C, et al. Increased viral shedding with aspirin treatment of rhinovirus infection. *JAMA*. 1975;231:1248-1251.
 138. Graham MH, Burrell CJ, Douglas RM, et al. Adverse effects of aspirin, acetaminophen, and ibuprofen on immune function, viral shedding, and clinical status in rhinovirus-infected volunteers. *J Infect Dis*. 1990;162:1277-1282.
 139. Dinarello CA. Endogenous pyrogens: the role of cytokines in the pathogenesis of fever. In: Mackowiak PA, ed. *Fever: Basic Mechanisms and Management*. New York, NY: Raven Press; 1991:23-47.
 140. Mellouk S, Green SJ, Nacy CA, Hoffman SL. IFN- γ inhibits development of *Plasmodium berghei* exo-

- erythrocytic stages in hepatocytes by an L-arginine-dependent effector mechanism. *J Immunol.* 1991;146:3971-3976.
141. Naotunne TDS, Karunaweera ND, Del Giudice G, et al. Cytokines kill malaria parasites during infection crisis: extracellular complementary factors are essential. *J Exp Med.* 1991;173:523-529.
 142. Curfs JHAJ, Van Der Meer JWM, Sauerwein RW, Eling WMC. Low dosages of interleukin 1 protect mice against lethal cerebral malaria. *J Exp Med.* 1990;172:1287-1291.
 143. Woodman JP, Dimier IH, Bout DT. Human endothelial cells are activated by IFN- γ to inhibit *Toxoplasma gondii* replication: inhibition is due to a different mechanism from that existing in mouse macrophages and human fibroblasts. *J Immunol.* 1991;147:2019-2023.
 144. Liew FY, Li Y, Millott S. Tumor necrosis factor α synergizes with IFN- γ in mediating killing of *Leishmania major* through the induction of nitric oxide. *J Immunol.* 1990;145:4306-4310.
 145. Torrico F, Heremans H, Rivera MT, Van Marck E, Billiau A, Carlier Y. Endogenous IFN- γ is required for resistance to acute *Trypanosoma cruzi* infection in mice. *J Immunol.* 1991;146:3626-3632.
 146. Ungar BVP, Kao T-C, Burris JA, Finkelman FD. Cryptosporidium infection in an adult mouse model: independent roles for IFN- γ and CD4⁺ T lymphocytes in protective immunity. *J Immunol.* 1991;147:1014-1022.
 147. Sambhi SK, Kohonen-Corish MRJ, Ramshaw IA. Local production of tumor necrosis factor encoded by recombinant vaccinia virus is effective in controlling viral replication in vivo. *Proc Natl Acad Sci U S A.* 1991;88:4025-4029.
 148. Feduchi E, Carrasco L. Mechanism of inhibition of HSV-1 replication by tumor necrosis factor and interferon. *Virology.* 1991;180:822-825.
 149. Strijp HAG, Van Der Tol ME, Miltenburgh LAM, Van Kessel KPM, Verhoff J. Tumor necrosis factor triggers granulocytes to internalize complement-coated virus particles. *Immunology.* 1991;73:77-82.
 150. Hedges S, Anderson P, Lidin-Janson G, Deman P, Svanborg C. Interleukin-6 response to deliberate colonization of the human urinary tract with gram-negative bacteria. *Infect Immun.* 1991;59:421-427.
 151. Vogels MTE, Vander Meer JWM. Use of immune modulators in nonspecific therapy of bacterial infections. *Antimicrob Agents Chemother.* 1992;36:1-5.
 152. Bernheim HA, Bodel T, Askenase PW, Atkins E. Effects of fever on host defense mechanisms after injection of the lizard *Dipsosaurus dorsalis*. *Br J Exp Pathol.* 1978;59:76-84.
 153. Dinarello CA. The proinflammatory cytokines interleukin-1 and tumor necrosis factor and treatment of the septic shock syndrome. *J Infect Dis.* 1991;163:1177-1184.
 154. Casey LC, Balk RA, Bone RC. Plasma cytokine and endotoxin levels correlate with survival in patients with the sepsis syndrome. *Ann Intern Med.* 1993;119:771-778.
 155. Johnson J, Brigham KL, Jesmok G, Meyrick B. Morphologic changes in lungs of anesthetized sheep following intravenous infusion of recombinant tumor necrosis factor α . *Am Rev Respir Dis.* 1991;144:179-186.
 156. Heinzel FP. The role of IFN- γ in the pathology of experimental endotoxemia. *J Immunol.* 1990;145:2920-2924.
 157. Henricson BE, Neta R, Vogel SN. An interleukin-1 receptor antagonist blocks lipopolysaccharide-induced colony-stimulating factor production and early endotoxin tolerance. *Infect Immun.* 1991;59:1188-1191.
 158. Ohlsson K, Björk P, Bergenfeldt M, Hageman R, Thompson RC. Interleukin-1 receptor antagonist reduces mortality from endotoxin shock. *Nature.* 1990;348:550-552.
 159. Opal SM, Cross AS, Sadoff JC, et al. Efficacy of antilipopolysaccharide and anti-tumor necrosis factor monoclonal antibodies in a neutropenic rat model of *Pseudomonas* sepsis. *J Clin Invest.* 1991;88:885-890.
 160. Overbeek BP, Veringa EM. Role of antibodies and antibiotics in aerobic gram-negative septicemia: possible synergism between antimicrobial treatment and immunotherapy. *Rev Infect Dis.* 1991;13:751-760.
 161. Alexander HR, Sheppard BC, Jensen JC, et al. Treatment with recombinant tumor necrosis factor-alpha protects rats against lethality, hypotension, and hypothermia of gram-negative sepsis. *J Clin Invest.* 1991;88:34-39.
 162. Fisher CJ Jr, Agosti JM, Opal SM, et al. Treatment of septic shock with tumor necrosis factor receptor: Fc fusion protein. *N Engl J Med.* 1996;334:1697-1702.
 163. Freudenberg MA, Galanos C. Tumor necrosis factor alpha mediates lethal activity of killed gram-negative and gram-positive bacteria in D-galactosamine-treated mice. *Infect Immun.* 1991;59:2110-2115.
 164. Gibson RL, Redding GJ, Henderson WR, Truog WE. Group B streptococcus induces tumor necrosis factor in neonatal piglets: effect of the tumor necrosis factor inhibitor pentoxifylline on hemodynamics and gas exchange. *Am Rev Respir Dis.* 1991;143:598-604.
 165. Birx DL, Redfield RR, Tencer K, Fowler A, Burke DS, Tosato G. Induction of interleukin-6 during human immunodeficiency virus infection. *Blood.* 1990;76:2303-2310.
 166. Radolf JD, Norgard MV, Brandt ME, Isaacs RD, Thompson PA, Beutler B. Lipoproteins of *Borrelia burgdorferi* and *Treponema pallidum* activate cachectin/tumor necrosis factor synthesis: analysis using a CAT reporter construct. *J Immunol.* 1991;147:1968-1974.
 167. Habicht GS, Katona LI, Benach JL. Cytokines and the pathogenesis of neuroborreliosis: *Borrelia burgdorferi* induces glioma cells to secrete interleukin-6. *J Infect Dis.* 1991;164:568-574.
 168. Jacobs RF, Jabor DR. The immunology of sepsis and meningitis-cytokine biology. *Scand J Infect Dis Suppl.* 1990;73:7-15.
 169. Jenkins JK, Carey PD, Byrne K, Sugerman, HJ, Fowler AA III. Sepsis-induced lung injury and the effects of ibuprofen pretreatment: analysis of early alveolar events via repetitive bronchoalveolar lavage. *Am Rev Respir Dis.* 1991;143:155-161.
 170. Saez-Llorens X, Jafara HS, Olsen KD, Nariuchi H, Hansen EJ, McCracken, GH. Induction of suppurative arthritis in rabbits by *Haemophilus* endotoxin, tumor necrosis factor- α , and interleukin-1 β . *J Infect Dis.* 1991;163:1267-1272.
 171. Rook GAW, Al Attiyah R. Cytokines and the Koch phenomenon. *Tubercle.* 1991;72:13-20.